

Antibacterial Activities of Perfume: Combination Flower *Magnolia alba*, *Cananga odorata* and *Mimusops elengi* L, Fixed with *Pogostemon cablin* Oil

Rosnani Nasution^{1*}, Azalia Izdhihar Azwar¹, Hira Helwati¹, Marianne²

¹Department of Chemistry, Faculty of Mathematics and Natural Sciences, Universitas Syiah Kuala, Banda Aceh 23111, Indonesia.

²Department of Pharmacology, Faculty of Pharmacy, Universitas Sumatera Utara, Medan 20155, Indonesia.

Abstract. The making of perfume from the combination *Magnolia alba* (*M. alba*) flower oil, *Cananga odorata* (*C. odorata*) and *Mimusops elengi* L (*M. elengi*) fixed with *Pogostemon cablin* oil (*P. cablin*) have been done. Essential oil of *M. alba* and *C. odorata* was distilled by water distillation and extraction essential oil of *M. elengi* flower using n-hexane evaporated solvent method. Composition perfume through organoleptic test obtained with a ratio of 4% v/v, 4% v/v, and 2% v/v for each essential oil *M. alba*, *C. odorata* and *M. elengi*. Antibacterial activity of the perfume and each of the volatile oil was performed by the disc diffusion method. Antibacterial activity test showed that perfumes were able to inhibit the growth of *Escherichia coli* (*E. coli*) and *Staphylococcus aureus* (*S. aureus*) bacteria, but less active. Antibacterial activity of the perfume was more active against *E. coli* with an inhibitory zone of 8.3 mm and 39.81% inhibitory power than *S. aureus* bacterial 7.3 mm inhibition zone with 37.55% inhibitory power. Perfumes after storage for one, and two months had better antibacterial activity than before storage with an average inhibitory zone of 11-15 mm and inhibitory power of 50-68,18%.

Keywords: Antibacterial; *E. coli*; *C. odorata*; *M. alba*; *M. elengi*; *P. cablin*; *S. aureus*

Abstrak. Pembuatan parfum dari kombinasi minyak bunga *Magnolia alba* (*M. alba*), *Cananga odorata* (*C. odorata*) dan *Mimusops elengi* L (*M. elengi*) yang difiksasi dengan minyak *Pogostemon cablin* (*P. cablin*) telah dilakukan. Minyak esensial *M. alba* dan *C. odorata* didistilasi dengan destilasi air dan ekstraksi minyak atsiri bunga *M. elengi* menggunakan metode pelarut n-heksana yang diuapkan. Komposisi parfum melalui uji organoleptik diperoleh dengan rasio 4% (v/v), 4% (v/v), dan 2% (v/v) untuk masing-masing minyak atsiri *M. alba*, *C. odorata* dan *M. elengi*. Aktivitas antibakteri parfum dari masing-masing minyak atsiri dilakukan dengan metode difusi cakram. Uji aktivitas antibakteri menunjukkan bahwa parfum mampu menghambat pertumbuhan bakteri *Escherichia coli* (*E. coli*) dan *Staphylococcus aureus* (*S. aureus*). Aktivitas antibakteri parfum lebih aktif terhadap *E. coli* dengan zona hambat 8,3 mm atau daya hambat 39,81% dibandingkan dengan bakteri *S. aureus* dengan zona hambat 7,3 mm atau daya hambat 37,55%. Parfum setelah penyimpanan selama satu dan dua bulan memiliki aktivitas antibakteri yang lebih baik berbanding sebelum penyimpanan, dengan zona hambat rata-rata 11-15 mm dan daya hambat 50-68, 18%.

Kata Kunci: Antibakteri; *E. coli*; *C. odorata*; *M. alba*; *M. elengi*; *P. cablin*; *S. Aureus*

Received 25 February 2019 | Revised 06 May 2019 | Accepted 08 May 2019

*Corresponding author at: Universitas Syiah Kuala, Banda Aceh, Indonesia.

E-mail address: rosnani@unsyiah.ac.id

1. Introduction

Perfume is a substance that has a fragrant and pleasant aroma, can be derived from plants, and can be a combination of various essential oils [1]. The use of essential oils is increasingly being developed in various perfume industries to reduce the negative impact of the use of synthetic substances and to provide aromatherapy effects [2]. However, the use of perfume that can function as an antimicrobial is still very lacking. Essential oils are organic compounds contained in plant parts [3], which contain monoterpenes and sesquiterpene compounds [4]. The distinctive aroma of essential oil has the potential to be developed into a natural scented perfume, the aroma of essential oil in the perfume is tied by adding fixative substances, one of the fixative substances that are often used is patchouli oil (*Pogostemon cablin*) [5].

Literature studies show that essential oils have the potential to be antimicrobial substances, drugs, pesticides, and cosmetics [6]. The content of monoterpenoid derivatives such as eugenol, thymol, and carvacrol in essential oils is thought to play an active role as an antimicrobial [7].

Some plants that contain essential oils, and are always used as perfumes are *M. alba* flowers, *M. elengi*, *C. odorata*, which have antimicrobial activities and other medicinal activities. In addition, *M. alba* plants also have pharmacological activity as antimicrobial, anti-cancer, anti-plasmodial, antioxidant and anti-depressant [8-9]. *M. elengi* plants have benefits as antibacterial, antifungal and anticancer [10]. *M. elengi* essential oil is efficacious as an ingredient of wound salts, boils, and leucorrhoea [11]. Flowers of *C. odorata* have antibacterial and antifungal activity [12]. The use of *C. odorata* as an aromatherapy plant effectively reduces depression, high blood pressure and anxiety [13].

Based on the data above, this study carried out, the combination of essential oils of flower from *M. alba*, *M. elengi* and *C. odorata* of which fixed using *P. cablin* oil, the perfumes hoped inhibits the growth of *E. coli* and *S. aureus* of microbes. Organoleptic tests by looking at the fragrance, preference, and sharpness of the scent through.

2. Methods

2.1 Plant Material

M. alba and *C. odorata* flowers taken in the village of Meunasah Timue, District Peusangan Selatan, Kab. Bireuen. *M. elengi* flowers are taken around the Universitas Syiah Kuala campus, Banda Aceh. Samples were taken in January 2017 and determined at the Biology Laboratory Herbarium Department of the Faculty of Mathematics and Natural Sciences, Universitas Syiah Kuala, Banda Aceh. Patchouli oil is obtained directly from the seller of patchouli essential oil in South Aceh. The bioindicator used in this study was the pure culture of *E. coli* and *S. aureus* bacteria obtained from the Microbiology Laboratory of the Faculty of Medicine, Universitas Syiah Kuala.

2.2 Phytochemical screening

The method used for testing of phytochemical can be found in: Phytochemical methods, A Guide to Modern Techniques of Plant Analysis [4]

2.3 Extraction of essential oils

Extraction of *M. alba* flower essential oil (4.7 Kg) and *C. odorata* (2.9 Kg) was carried out by the water distillation method. Each fresh flower is sliced and distilled water. Distillation was carried out for 5-8 hours [14]. Extraction of *M. elengi* flower essential oil was carried out by evaporated solvent extraction method using *n*-hexane solvent.

M. elengi flowers fresh was dried for three days to remove water in flowers, mashed and macerated using *n*-hexane for 3 x 24 hours. The filtrate produced is evaporated with a rotary evaporator at a temperature below 35°C to obtain concrete essential oil.

2.4 Refractive index

Determination of the refractive index of each *M. alba*, *M. elengi*, and *C. odorata* essential oil of flowers was carried out with a refractometer [15].

2.5 The making of perfume

Perfume making is made with a combination of three essential oils based on organoleptic tests. Organoleptic tests were carried out by measuring the subjective attitude of respondents based on organoleptic characteristics. The data provided is in the form of a personal response questionnaire about the level of fragrance, favorite and sharpness of a combination of perfume. This test uses respondents and there is no comparison test material. The test subjects use a scale of 1-5 [16]. Respondents consisted of 40 people with age criteria (20-25 years). The form of the organoleptic test form can be seen in Table 1.

Table 1. Forms of organoleptic combinations of three essential oils

No.	Fragrance Level	Scale Favorite Level	Sharpness Level
1	Very fragrant	Very like	Very sharp
2	Fragrant	Likes	Sharpness
3	A little fragrant	Little likes	A little sharp
4	Not fragrant	Don't like	Not sharp
5	Very not fragrant	Very dislike	Very not sharp

The combination of essential oils was carried out with variations in composition by looking at the effect of one variable on the other variables based on the percentage of use of each essential oil. Then the optimum composition test for *M. Alba* oil was carried out, tested the optimum

composition for essential oils of *C. odorata*, tested the optimum composition for *M. Elengi* essential oil, so that the optimum composition of each essential oil (in percent) was then used as a constituent of perfume which is found in Table 2.

Table 2. The composition of essential oils making up perfume (combination of three essential oils)

Composition (% b/v)				
<i>M. alba</i>	<i>C. odorata</i>	<i>M. elengi</i>	<i>P. cablin</i>	Alcohol
4	4	2	5	make up to 100

2.6 Antibacterial activity test

Antibacterial activity of perfume and each essential oil against *E. coli* and *S. aureus* were carried out by disc diffusion method. Antibacterial testing is carried out by placing disc paper that has been filled with perfume and essential oil on the test media which has been inoculated with *E. coli* and *S. aureus* bacteria respectively. The positive control of the antibacterial test was gentamicin, while the alcohol and *n*-heksana were negative controls for each test. Incubation is carried out for 18 hours to see the inhibitory zone against antibacterial. The inhibition zone is measured using a ruler in millimeters [17]

3. Result And Discussion

3.1 Essential oil extraction

Fresh flower distillation of *M. alba* (4.7 Kg) and *C. odorata* (2.9 Kg) was carried out for 4-8 hours [14]. The results obtained from *M. alba* flower was 5.98 mL of essential oil with a yield of 0.1% (v / b), and from *C. odorata* flowers obtained 8.07 mL of essential oil with a yield of 0.2% (v / b). The extraction of *M. elengi* flower essential oil using the *n*-hexane vaporizing solvent method yielded a yield of 0.714% (b / b).

The results of essential oils obtained from the three flowers above are tested their phytochemical, and the refractive index is tested.

Phytochemical test results on fresh samples and essential oils of *M. elengi* showed terpenoids. The terpenoid test was carried out using the Liebermann-Burchard reagent, this test showed positive results with observations of the formation of a red color[4]

3.2 Refractive index

The test results of the refractive index values of the essential oils of *C. odorata*, *M. alba*, and *M. elengi* are shown in Table 3.

Table 3. Essential oil refraction index test

Essential oil	Refraction index (nD)
<i>C. odorata</i>	1,483
<i>M. alba</i>	1,455
<i>M. elengi</i>	1,488

The three essential oils above meet the value of the refractive index of essential oils in general. There are several essential oils that already have quality standards (SNI), one of which is Cananga oil or Essential oil of *C. odorata*. Based on SNI quality standards the refractive index of *C. odorata* oil ranges between (1493-1503).

3.3 The Perfume Organoleptic

Organoleptic testing is carried out including the test of fragrance level, aroma preference, and sharpness. This aims to determine the optimum composition of each constituent oil as a perfume. The optimum composition of each essential oil was determined from the percentage level of the panelists with the criteria of fragrant, like and have a non-sharp aroma. Analysis of the panelists response to the variation in the concentration of essential oil of *M. alba* is shown in Figure 1. Analysis of the panelists response to the concentration of *C. odorata* essential oil is shown in Figure 2 and analysis of the response of panelists to the concentration of *M. elengi* essential oil is shown in Figure 3.

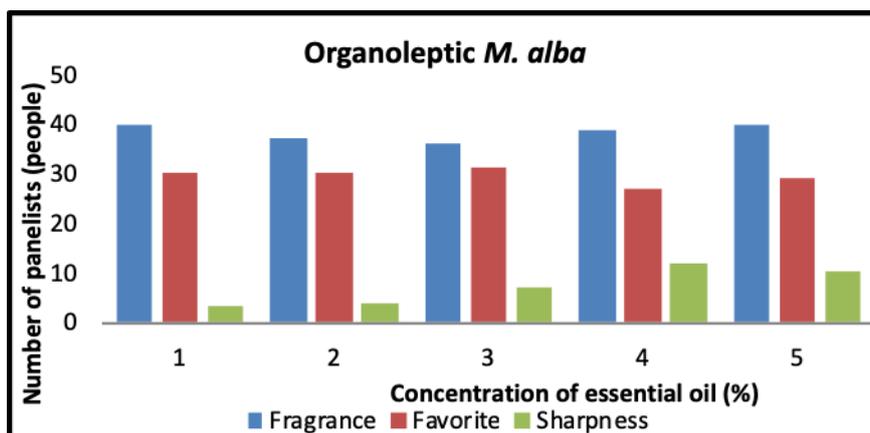


Figure 1. Respond from panelists to variations in the concentration of essential oils *M. alba* flower

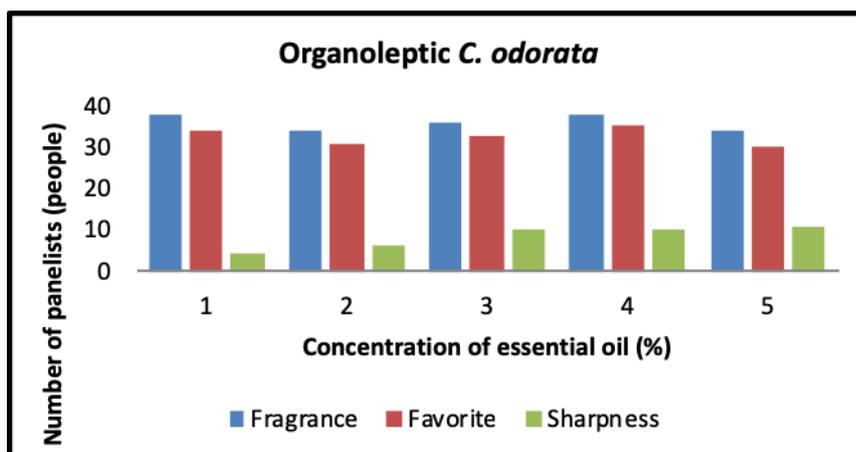


Figure 2. Respond from Panelists to variations in the concentration of essential oils *C. odorata* flower

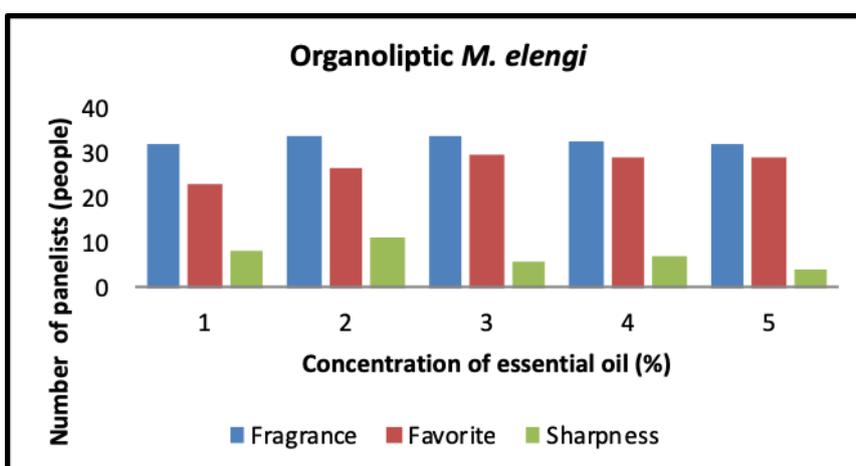


Figure 3. Respond from panelists to variations in the concentration of essential oils *M. elengi* flower

Based on figures 1, 2, and 3 above, it shows that the optimum composition is 4% (v / v) essential oil of *M. alba* flower, 4% (v / v) essential oil *C. odorata* and 2% (v / b) essential oil concrete *M. elengi* . The results of the organoleptic data statistic analysis were performed using the T-test by looking at the best concentration ratio. Perfume with the optimum composition of organoleptic test results (the most preferred by panelists) was tested for antibacterial activity, against *S. aureus* and *E. coli* bacteria.

3.4 Antibacterial Activity

The results of the antibacterial activity test of each essential oil, a combination of essential oils and essential oils in alcohol are shown in Table 4.

Table 4. Inhibition zones (mm) of each essential oil, perfume (combination of three essential oils), each essential oil with alcohol, against *S. aureus* and *E. coli* bacteria

Essential oil	Inhibitory zone (mm)	
	<i>S. aureus</i>	<i>E. coli</i>
<i>P. cablin</i>	12,6	12,6
<i>C. odorata</i>	13,6	11,6
<i>M. elengi</i>	6	7,3
<i>M. alba</i>	13	13
Essential oils with the addition of alcohol		
2% <i>M. elengi</i>	8,6	8
4% <i>C. odorata</i>	8,6	8,3
4% <i>M. alba</i>	8,3	7,6
5% <i>P. cablin</i>	10,3	11
Combination of essential oils		
<i>P. cablin</i> + <i>C. odorata</i> + <i>M. alba</i> + <i>M. elengi</i> + alcohol	8,6	8,3
<i>P. cablin</i> + <i>C. odorata</i> + <i>M. alba</i> + alcohol	13	11
<i>P. cablin</i> + <i>M. alba</i> + <i>M. elengi</i> + alcohol	7,6	8
<i>P. cablin</i> + <i>C. odorata</i> + <i>M. elengi</i> + alcohol	10	8,6

Key: disc diameter (6 mm)

Based on data on the diameter of the inhibition zone (mm) in Table 4 above, it can be seen that each essential oil and perfume can inhibit the growth of *S. aureus* and *E. coli* bacteria. *C. odorata* essential oil was able to inhibit the best growth of *S. aureus* bacteria with an inhibition zone of 13.5 mm and obtained a percentage of activity of 59.13%, this value was said to actively inhibit the growth of *S. aureus* bacteria. This is in line with the research conducted [18], showed that essential oil of *C. odorata* was able to inhibit bacterial growth with an inhibition zone of 12-24 mm.

M. alba essential oil has the best activity to inhibit the growth of *E. coli* bacteria with an inhibition zone of 13 mm and a percentage of activeness of 63.10% so that it can be said that *M. alba* essential oil actively inhibits the growth of *E. coli* bacteria. This is in line with the results of a study conducted [18] proved that *M. alba* essential oil was able to inhibit the growth of Gram-positive and Gram-negative bacteria with an average inhibition zone of 11-17 mm.

The antibacterial activity of *M. elengi* essential oil was able to inhibit the growth of *S. aureus* bacteria with an inhibitory zone of 6 mm, while the inhibition zone of *E. coli* bacteria was 7.3 mm. When compared with the inhibitory activity of the growth of *S. aureus* essential oil, *M. elengi* is more able to inhibit the growth of *E. coli* bacteria which is a Gram-negative bacterium, this result is in line with that reported [19] that *M. elengi* flower essential oil is only active against Gram-negative bacteria.

The antibacterial activity of each essential oil added to alcohol was obtained by a relatively small inhibition zone compared to the pure essential oil activity, except for *M. elengi* essential oil with a concentration of 2% having better activity than the pure concrete essential oil. Patchouli essential oil with a concentration of 5% decreased slightly compared to pure oil.

Perfume (a combination of three flower essential oils namely *M. alba*, *C. odorata*, and *M. Elengi*) which were fixed with *P. cablin* oil was able to inhibit the growth of *S. aureus* bacteria with an inhibitory zone of 8.6 mm with a percentage of activity of 39.81 %. Based on the percent value of activity obtained, it can be said that perfume which is fixed with *P. cablin* oil is less active to inhibit the growth of *S. aureus* bacteria. The test results of perfume activity on the growth of *E. coli* bacteria obtained an inhibition zone of 8.3 mm with a percentage of activity of 37.55%. Perfume activity that is less active against these bacteria, is thought to be due to a reaction between the active groups present in each essential oil so as to reduce the activity of a compound. In addition, it is also suspected that the effect of the irregularity of *M. elengi* essential oil can influence antibacterial activity.

According to the literature, *C. odorata* essential oil contains caryophyllene (36.44%) and linalool (5.97%) compounds which are active as antibacterial [20] and in *M. alba* flower essential oil there is linalool (85.78%) and caryophyllene (0.92%) (Punjee et al., 2009). [14]

The antibacterial mechanism of linalool and caryophyllene is the same, by damaging the bacterial cytoplasmic membrane and coagulating cell components [21]

Antibacterial activity is also carried out on perfume after experiencing a storage period of one and two months. Perfume data with the effect of storage period are shown in (Table 5).

Table 5. The zone of inhibition (mm) of perfume after storage of one and two months

Composition	Inhibitory zone (mm)	
	<i>S.aureus</i>	<i>E.coli</i>
Alcohol	8	8,5
<i>n</i> -hexane	-	-
One month storage perfume	11	11,5
Two months of storage perfume	13,5	15
Gentamicin 10µg/mL	22	22

Key: disc diameter (6mm)

Inhibitory zone against *E. coli* and *S. aureus* are shown in Figure 4.

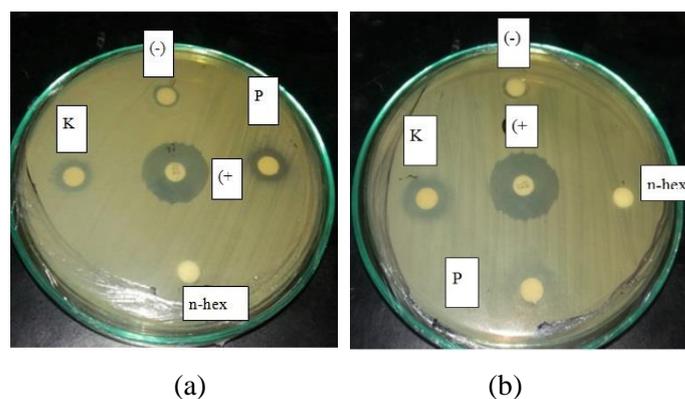


Figure 4 a. Perfume inhibition zone with storage time variation for *E. coli* bacteria; b. Perfume inhibition zone with storage time variation for *S. aureus* bacteria

The perfume storage period carried out in this study was able to influence antibacterial activity. This is evidenced by a better perfume inhibition zone and a higher percentage of resistance after storage compared to the inhibition zone before storage. After one month of storage, the inhibition zone of perfume for *S. aureus* bacteria was 11 mm and after storage for two months, an inhibition zone of 13.5 mm was obtained when compared to the positive control inhibitory zone of 22 mm, the percentage of activity was 50% for one month storage period, and 61, 36% for two months storage, this value can be said to actively inhibit the growth of *S. aureus* bacteria

The increase in the inhibition zone of perfume was also obtained in antibacterial activity against *E. coli* bacteria with an inhibition zone of 11.5 mm for one-month storage and 15 mm for two months of storage. Compared to the positive control of 22 mm, the percent of activity was 52.27% for one-month storage and 68.18% for storage for two months. The percent value of activity obtained is said to actively inhibit the growth of *E. coli* bacteria. Increased activeness of perfume after storage is suspected during the storage period of each active component able to combine so as to form a synergistic activity in inhibiting the growth of the two test bacteria in addition to the influence of essential oil of *P. cablin* after one month can increase the activity of perfume. However, this research needs to be carried out further. In general, essential oils and perfumes combined with essential oils of *M. alba*, *C. odorata*, and *M. elengi* are broad-spectrum antibacterial substances. Because it can inhibit bacterial growth from gram positive and gram negative

The monoterpenes group in essential oil acts as an antibacterial which reacts with porin (transmembrane protein) in the outer membrane of the bacterial cell wall, forming a strong polymeric bond that causes damage to the porin. When the porin has been damaged it will reduce

the permeability of the bacterial cell wall which will cause bacterial cells to lack nutrients, so that bacterial growth is inhibited or dies [22]. In addition, essential oils that are active as antibacterial generally contain groups (-OH) and carbonyl. Bacteriostatic activities of each essential oil and perfume are generally more able to inhibit the growth of *S. aureus* bacteria than *E. coli* bacteria. this is because the *S. aureus* bacteria are gram-positive bacteria that have a simpler composition of cell walls compared to *E. coli* bacteria which are gram-negative bacteria. [23], reported that gram-negative bacteria had a higher lipid composition compared to gram-positive bacteria. Generally, gram-negative bacteria produce mucus as a form of self-defense from chemical compounds that harm it. The outer membrane of gram-negative bacteria acts as a barrier to the entry of compounds that are not needed by cells, such as bacteriocins, enzymes, and compounds that are hydrophobic [24]. Antimicrobial compounds work by being able to penetrate lipopolysaccharides (LPS) from the cell wall. Hydrophilic molecules are easier to pass through LPS compared to hydrophobic ones. Gram- positive bacteria do not have LPS, so nothing prevents the entry of antibacterial substances and molecules of antimicrobial compounds that are hydrophilic and hydrophobic (such as essential oils) can diffuse into cells easily [25].

4. Conclusion

Based on the results of research that has been carried out, it can be concluded that the test for antibacterial perfume activity (a combination of *M. alba* essential oil, *C. odorata*, and *M. elengi*) which is fixed with *P. cablin* oil capable of inhibiting the growth of Gram-negative and Gram-positive bacteria with an inhibition zone of 8.3-8.6 mm with a percentage of activity (inhibitory power) of 39.71-38.56%. The period of storage of perfume (in this study) can affect antibacterial activity, after one month of storage, the inhibition zone of perfume for *S. aureus* bacteria was 11 mm and after storage for two months for *S. aureus* bacteria, an inhibition zone of 13.5 mm, percentage of activity (inhibitory power) was 50%, and 61, 36% (compared to Positive control), while for *E. coli* bacteria was 52.27% for one-month storage and 61.36% for two month storage.

REFERENCES

- [1] Webster. M, “*Definition Of Perfume*”, 2017. [Online]. Available : <https://www.merriam-webster.com/dictionary/perfume>. [Accessed: July. 31st, 2017].
- [2] Aldo, “Penetapan Kadar Benzaldehid pada Sampel Parfum “X” dari 3666 Toko Parfum di Wilayah Surabaya Selatan,” *Jurnal Ilmiah Mahasiswa*, no. 4, pp. 1-11. 2015.
- [3] N.S. Rupilu, Y.F. Lamapaha, *Potensi Lengkuas Sebagai Antimikroba (Studi In Vitro pada Gram Negatif)*. Universitas Negeri Malang, Malang: 2008.
- [4] Harborne. J.B. *Phytochemical Methods: A Guide to Modern Techniques of Plant Analysis, 3rd Edn.*, Chapman and Hall, London, p.302, 1998.
- [5] I. Mustika dan I. Nuryani, “Strategi Pengendalian Nematoda Parasit pada Tumbuhan Nilam,” *Jurnal Litbang Pertanian XXV*, vol. 4, no. 1, pp.7-15, 2006.
- [6] S.Y. Hartati, “Prospek Pengembangan Minyak Atsiri Sebagai Pestisida Nabati”, *Jurnal Perspektif*, Vol. 11, No 1, pp.45-58, 2012.

- [7] M.B. Isman, "Plant Essential Oils For Pets and Disease Management", *Crop Protection*, Vol. 19, No. 8, pp.603-608, 2000.
- [8] S.W. Lee, W. Wee, Y.Y.F Siong, D.F. Syamsunir, "Characeristic of Antimicrobial, Antioxidant, Anticancer Property and Chemical Composition of Michelia alba Seeds and Flower Extract", *Stamford Journal of Pharmaceutical Sciences*, 2014.
- [9] H. Tan-li, H. Ming-wu, H. Liang-chen, C. Ming-liu, and C. Yi-chen, "The Pharmacological Activities of (-) - Anonaine" *Molecules Journal*, Vol. 18, pp.8257-8263, 2013.
- [10] M.S Baliga, R.J Pai, H.P Bhat, P.L Paliatty, R. Bloor, "Chemistry and Medicinal Properties of The Bakul (*Mimusops elengi* Linn) : A Review," *Food Research Internasional*, Vol. 44, Issue. 7, pp.1823-1829, 2011.
- [11] M. Roqaiya, W. Begum, S.F. Majeedi and A.Sayed, "A Review on Traditional Uses and phytochemical Properties of *Mimusops elengi* Linn ", *International Journal of Herbal Medicine*, Vol. 2, No. 6, pp.20-23, 2015.
- [12] N. Zaine, *Volatile Compounds and Biological of extract of Curanga odorata and It's petal – Derived Cells*, University Putra Malaysia, Malaysia, 2009.
- [13] L.T Tan, L.H. Lee, W.F. Yin, Ck. Chan, H. Abdul Kadir, K.G Chan, B.H. Goh, "Traditionl Uses, Phytochemistry and Bioactivities of *Cananga odorata* (Ylang-ylang)", *Hindawi Publishing Corporation, Evidence-Based Complementary and Alternative Medicine*, Vol. 2015, Article ID: 896314.
- [14] P. Punjee, U. Dilokkunanant, U. Sukatta and S. Vajrodaya, "Scented Extracts and Essential Oil Extraction from *Michelia alba* D.C", *Kasetart journal (Nat-Sci)*, Vol. 43, pp.197-203, 2009.
- [15] F.P Iswara, D. Rubiyano dan T.S Julianto, "Anlisis Senyawa Berbahaya dalam Parfum dengan Kromatografi Gas-Spektrofotometri Massa Berdasarkan Material Safety Data Sheet (MSDS)", *Indo Journal of Pharmaceutical Research*, Vol. 1, Issue 2, pp.18-27, 2014.
- [16] Indonesia, Direktorat Pendidikan Dasar dan Menengah Kejuruan, Departemen Pendidikan Nasional, "Menguji Kesukaan secara Organoleptik", Jakarta : 2003.[Online]. Available : http://psbtik.smkn1cms.net/pertanian/agroindustri/agroindustri_non_pangan/menguji_kesukaan_secara_organoleptik.pdf
- [17] B.W Lay, *Analisis Mikrobiologi di Laboratorium*, Raja Grafindo, Jakarta, 1994.
- [18] K.S. Sree, M. Anudeep, Ch. Bhaskar, "Screening of Antimicrobial Activity of Flower Extracts on Human Bacteria Pathogens", *Journal of Pharmacognosy and Phytochemistry*, Vol. 3, No. 6, pp.153-156, 2015.
- [19] A. Faheem, C.K. Wong, I.M Eldeen, M.Z. Asmawi, H. Osman, "Volatile and Bioactivities of *Mimusops elengi* Linn Flowers", *Latin American Journal of Pharmacy*, Vol. 31, No. 2, pp.331-335, 2012.
- [20] R. Pujiarti, T.B. Widowati, K. Kasmudjo, S. Sunarta, "Kualitas, Komposisi Kimia dan Aktivitas Antioksidan Minyak Kenanga *Cananga odorata*", *Journal of Forest Science*, Vol. 9, No. 1, 2015.
- [21] S. Burt, "Essentils oils : Their Antibacterial Properties and Potential Applications in foods", *International Journal of Food Microbiology*, Vol. 94, Issue. 3, pp 223-253, 2004.
- [22] Cowan, "Plant Products as Antimicrobial Agents", *Clinical Microbiology Review*, Vol. 12, No. 4, pp.564-582, 1999.
- [23] E. Jawetz, J.L Melnick and E.A Adelberg, *Mikrobiologi Kedokteran*, Salemba Medika, Jakarta, 2001.
- [24] P. M Davidson, J.N Sofos and A.L Branen, *Antimicrobial in Food, 3rd edition.*, Taylor and Francis Group, London, 2005.
- [25] M. Oussalah, S. Callet, L. Saucier and M. Lacroix, "Antimicrobial Effects of Selected Plant Essential Oils on the Growth of a *Pseudomonas putida* strain Isolated from Meat," *Meat Science*, Vol. 73, Issue. 2, pp 236-244, 2006.